Instruction of the Clean-up Process Using SENSIColumn IAC Aflatoxin 3ml Product Code: EFAF3105



Clean-up of Commodity Extracts of Food and Feed Samples containing Aflatoxins Via Immunoaffinity Chromatography and Subsequent HPLC Analysis with Postcolumn Derivatization

Subsequent HPLC Analysis with Postcolumn Derivatization using Pyridinium Perbromide

Principle:

This instruction of Aflatoxin total determination in food and feed focuses on the enrichment step of extract using immunoaffinity column (IAC) and quantification with HPLC.

Accepted laboratory extraction methods could be maintained. Full performance of the IAC column is given if pronounced criteria of organic solvent tolerance, elution process of analyte and working range of column is followed.

Many pretreatment methods of aflatoxin (AFG₂, AFG₁, AFB₂ and AFB₁) determination in food and feed show low sensitivity because of interfering substances if problematic matrices are applied.

This method of content determination of aflatoxins combines the high selectivity of an immunoaffinity column (IAC) with its potential to concentrate eluate and additional step of purification by HPLC column.

Please notice that this instruction focuses on the <u>handling with the IAC column</u>. For the commodity extraction step a literature method is given. The given apparatus (e.g. HPLC system) might serve as example among other possibilities.

Extraction (Literature method given):

Assuming that 25g sample are extracted by a total of 100ml methanol/water (60/40 v/v). If organic solvent proportion is varied the dilution of extract with PBS should be adapted accordingly in the enrichment step. On the other hand, if proportion of sample quantity and volume of extraction solvent is altered, calculation of gram equivalents must be corrected.

For groundnut meal as an example of a problematic matrix, literature method of Roch et al.¹ could be applied.

Enrichment Step IAC:

4ml extract (see above, <u>contains the quantity of aflatoxins of 1g sample</u>) are diluted with 16ml 50mM PBS (pH=7.4) and then applied in a reservoir on top of the **SENSIC** *olumn* **IAC Aflatoxin** *3ml* column.

To maintain full performance of the column, please take care that proportion of dilution buffer in the solution on top of the column is not too small. A proportion of 12% methanol, resulting in this example enrichment, does not affect column performance.

<u>Caution:</u> The proportion of organic solvent of PBS diluted extract, which is applied on the column, should not exceed 20% methanol and 10% acetonitrile.

If organic solvent proportion lies above these limits, recovery rates are diminished. Increase of diluted extract volume by diluting extract with additional PBS, on the other hand, has almost no consequences to column performance.

If samples are to be prepared simultaneously, manifold of J.T. Baker for 12 samples has proven of value. Rate of flow through the affinity gel is 1 to 3 ml/min. In case of problematic matrices rate of flow should lie below 2ml/min.

<u>Caution!</u> Be aware that no big air bubbles are neither in the gel nor between gel and luer lock outlet of column which prevent a permanent flow or necessary exchange of matter.

Depending on application and on expected contents, larger or smaller extract aliquots can be applied. In such cases the sample calculation (see below) must be adapted. **Wash:**

After whole sample has passed through the gel the latter is washed with 5ml of 10mM PBS/Methanol (90/10 v/v). Remaining liquids in the gel are removed by applying either pressure from top of the column or underpressure from bottom.

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Elution:

Sample reservoir on top of the **SENSI***Column* IAC Aflatoxin *3ml* column is removed and an appropriate vial is placed below the affinity column. The bound toxins are eluted by using a total of <u>2ml</u> of methanol as elution solvent. The elution process is performed in two steps to ensure complete release of analytes. First, a volume of 1ml elution solvent is applied. After that volume has passed through column half a minute is waited before the second portion of 1ml of elution solvent is eluted through the column. The flow rate of the elution process should not exceed 1ml/min very much. Remaining solvent solutions should be eluted by application of slight under- or overpressure. All methanolic fractions are unified to give the column eluate.

The column eluate may be injected into the HPLC directly or in case contents are low (<2ng/g, related to commodity content), it may be concentrated by evaporation (e.g. using VLM evaporator at 50°C under a permanent stream of nitrogen).

<u>Caution:</u> If you follow concentration by evaporation, add 100µl of acetic acid/water (50/50 v/v) as keeper. To avoid loss of analytes, stop concentration at a small volume of residue, e.g. 50 to 100µl.

The residue then is redissolved in HPLC solvent (e.g. 0.4ml) and an aliquot is finally injected into the system.

IAC Column Characteristics:

A) Working Range and Recovery Rates of SENSIColumn IAC Aflatoxin 3ml:

Working Range of Column: Zero Contamination of Column:	0.04 –200ng Aflatoxin total per IAC <0.04ng (LOD of HPLC-FLD method)					
Guaranteed Recovery Rates ^(*) within the Working						
Range:						
Aflatoxin total:	>90% ± 5%					
Aflatoxin B_1 (AFB ₁):	>95% ± 5%					
Aflatoxin B_2 (AFB ₂):	>90%% ± 5%					
Aflatoxin G_1 (AFG ₁):	>95%% ± 5%					
Aflatoxin G ₂ (AFG ₂):	>90% ± 5%					
⁽¹⁾ Becovery rates are confined to the complete IAC separation procedure (not including recovery r						

Recovery rates are confined to the complete IAC separation procedure (not including recovery rates of commodity extraction process), that means for the IAC Enrichment Step, organic solvent content of diluted extract below 20% methanol or 10% acetonitrile, for the Wash and Elution Step in the manner as described in this instruction

B) Cross Reactivities^(**) of SENSIColumn IAC Aflatoxin 3ml Column:

Aflatoxin B_1 (AFB ₁):			100%			
Aflatoxin B_2 (AFB ₂):			86%			
Aflatoxin G_1 (AF G_1):			101%			
Aflatoxin G_2 (AFG ₂):			82%			
(**) Recovery rate of AEB	AEC.	AEG	divided by recovery rate of AEB	if a total of 1	8110	Aflatovi

Recovery rate of AFB₂, AFG₁, AFG₂, divided by recovery rate of AFB₁ if a total of 1.8µg Aflatoxin total (with molar ratio of B₁, B₂, G₁, G₂, = 4:1:4:1) is analysed per column. Please notice that this quantity is near the capacity limit of column where binding sites of column are limited. For this reason cross reactivities of AFG₁ and AFG₂ are much higher at aflatoxin concentrations within the given working range where antibody binding sites are in excess (see table A above).

C) Capacity^(***) of SENSIColumn IAC Aflatoxin 3ml Column:

Maximum Column Capacity:	1.8µg Aflatoxin total
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An excess of aflatoxins, e.g. 4µg, in a small volume of 2ml PBS is incubated with the IAC for 5 minutes; then the IAC is washed with 2ml PBS and the nonbonded fraction is analyzed. The difference of added analyte and nonbonded analyt equals maximum column capacity.

Analytical Method:

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<u>HPLC:</u> Shimadzu; <u>Column:</u> Trentec Reprosil-Pur RP C18 120 ODS3 5µm; 125x3,0mm with guard column; <u>Mobile Phase A:</u> methanol / deionized water (85/15 v/v); <u>Mobile Phase B:</u> methanol / acetonitrile / deionized water (18/18/64 v/v/v); <u>Gradient:</u> 0.01 min B 100 %; 16 min B 100 %; 17 min B 0 %; 19 min B 0 %; 20 min B 100 %; <u>Flow Rate:</u> 0.5ml/min; <u>Time of Analysis</u>: 30min; <u>Injector Volume:</u> 100µl

<u>Post Column Derivatization</u>: 32 ppm pyridinium hydrobromide perbromide in dioxan/ deionized water (0.1/99.9, v/v); <u>Flow Rate</u>: 0.2ml/min pumped in reaction capillary which is put between end of column and detector by tee-connector; <u>Reaction Capillary</u>: PEEK-capillary, 1/16^{\prime} x 0.25 mm ID; length: 40 cm; <u>Tee-connector</u>: Tee column end fitting 1/16" x 1/16" x 1/16", 0.25mm in/out, 0.25mm side. <u>Fluorescence-Detection</u>: λ_{EX} [nm]: 362nm; λ_{EM} [nm]: 440nm.

Temperatur: Machine and eluents are at room temperature. Eluents are degassed with helium gas.

HPLC Method Characteristics

Aflatoxin B₁

Measuring range is linear from 10pg to 500pg Aflatoxin B₁ per injection (R^2 =0.999) in given HPLC method. Lower limit of detection (LOD) is 2pg AFB₁ per injection (signal to noise (S/N) ratio = 3).

Balance of commodity contamination and HPLC measuring range:

If stated dilution steps (enrichment step, eluate concentration step) are followed, aflatoxin B_1 contents in commodities of 0.04 to 2ng/g lie within measuring range of HPLC method. If contents are higher, from 2 to 200ng/g, IAC column eluate should be diluted accordingly with HPLC eluent or, alternatively, injector volume should be adjusted.

Only if commodity contents lie above working range of IAC column of 200ng/g total aflatoxin, the 1g equivalent per column of this instruction must be lowered. Thus, the IAC enrichment step and subsequent HPLC analysis should be repeated with a smaller extract volume, e.g. instead of 4ml extract a volume of 2ml extract, or even less where applicable, are diluted with 10ml PBS and applied to the IAC column.

Left Aflatoxin G₁, Aflatoxin B₂; Aflatoxin G₂:

Sensitivities [AREA Peak / pg] of aflatoxins AFG_1 , AFB_2 und AFG_2 related to that of AFB_1 are 37%; 136% and 67%, respectively.

AFG₁: Measuring range is linear from 25pg to 500pg Aflatoxin G₁ per injection (R^2 =0.999). LOD of detection is 6pg AFG₁ per injection (signal to noise (S/N) ratio = 3).

AFB₂: Measuring range is linear from 2.5pg to 125pg Aflatoxin B₂ per injection (R^2 =0.999). LOD is 1pg AFB₂ per injection (signal to noise (S/N) ratio = 3).

AFG₂: Measuring range is linear from 2.5pg to 125pg Aflatoxin G₂ per injection (R^2 =0.999). LOD is 2pg AFG₂ per injection (signal to noise (S/N) ratio = 3).

Because left aflatoxins (AFG₁, AFB₂ and AFG₂) mostly occur in common and are thus analyzed simultaneously with key aflatoxin B₁, different levels of commodity contents and established HPLC measuring ranges are balanced by the same manner as stated for aflatoxin B₁.

Example Sample Calculation of AFB₁ content:

(Calculation of AFB₂, AFG₁ and AFG₂ content is analogous)

A) Calculation of Sample Gramm Equivalents per HPLC injection:

25g Sample	х	4ml Extract	0.1ml x injector	0.1ml injector	=	0.25g Sample
100ml Extraction Solvent		0.4ml		volume		Equivalents

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B) Calculation of Aflatoxin B1 contamination of examined commodity in ng/g:

ng injected AFB₁ Sample Equivalents [g]

ng/g AFB₁ in e.g. ground nut meal

Buffer, Chemicals, Apparatus and Literature:

Phosphate Buffered Saline pH 7.4 (= 50mM PBS)

1.24g KH₂PO₄ 7.27g K₂HPO₄ 8.76g NaCl **Chemicals:** •acetonitrile, HPLC grade •methanol, HPLC grade •acetic acid, 100% ultrapure deionized water dipotassium hydrogenphosphate, >98% potassium dihydrogenphosphate, >98% sodium chloride

Evaporation:

•nitrogen gas 5.0 [Air Liquide M55763810] (to evaporate IAC-eluate)

Postcolumn Derivatization:

•pyridine hydrobromide perbromide, >95%. •dioxane, 99.5%

Apparatus:

HPLC; Shimadzu; pump: LC-6A (2 pieces); auto sampler: SIL 6B; fluorescence detector: RF-10AXL; data handling: CLASS LC10

Evaporator (with tripod) [VLM EVA EC1-S]

Vacuum SPE Manifold (BAKER spe-24G Column Processor - process up to 24 samples) [J.T. Baker 7208]

Dissolve in 1L deionized water. If necessary adjust pH to 7.4 (± 0.3) with 1N NaOH or 1N HCl

Consumables:

SENSIColumn IAC Aflatoxin 3ml

Elution Solvent:

methanol

Keeper:

acetic acid/water (50/50 v/v): Mix 10ml acetic acid and 10ml deionized water

Reagent:

32 ppm pyridine hydrobromide perbromide in dioxan / deionized water (0.1/99.9, v/v):

Partially dissolve 32mg pyridine hydrobromide perbromide in 1ml dioxane and pipet the oily suspended liquid into 1L degassed deionized water

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¹ "Determination of aflatoxins in groundnut meal by high-performance liquid chromatography: a comparison of two methods of derivatisation of aflatoxin B₁" G. Roch, G. Blunden, J. Haig, D.Coker and C. Gay B. J. Biomed Sci 1995, 52, 312-316