

VIRSeek Food Hepatitis E Virus

TEST KIT FOR HEPATITIS E VIRUS (HEV)
QUALITATIVE REAL-TIME RT-PCR FROM FOOD SAMPLES

Cat. No. Katalognummer 5728200501

For 48 real-time RT-PCR reactions

VIRSeek

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1 INTRODUCTION

Hepatitis E is a viral liver disease caused by hepatitis E virus (HEV) that spreads mainly via the faecal-oral route through person-to-person contact or contaminated food and water. In Europe, mainly HEV genotype 3 is linked to food-borne transmissions; thereby pigs and wild boars seem to be the main source of the virus. There is increasing evidence that the infection may be linked to undercooked pork and pork products. Usually the infection is self-limiting and resolves within 2–6 weeks. Occasionally a serious disease, known as fulminant hepatitis (acute liver failure) develops and a proportion of people with this disease can die. Safe water supply and food safety are amongst the most effective ways to combat the disease and are therefore key to the prevention of outbreaks. An effective vaccine to prevent hepatitis E virus infection has been developed and licensed in China, but it is not available elsewhere.

HEV is a non-enveloped single-stranded RNA virus. The virus is known to persist in the environment and on food surfaces comparable to other non-enveloped small RNA viruses. Additionally it has been shown that it can tolerate food-production processes which are routinely applied to control common bacterial pathogens.

The VIRSeek Food Hepatitis E Virus kit provides all reagents for the rapid detection of the hepatitis E virus in a number of food products including soft fruits, leaf, stem and bulb vegetables, bivalve molluscan shellfish as well as pork liver containing processed meat products such as sausage. A viral concentration prior to RNA extraction is required for these sample types.

The VIRSeek Food Hepatitis E Virus kit is validated for use with the Agilent AriaMx™, Bio-Rad CFX96 Touch™ and CFX96 Touch™ Deep Well PCR platforms.

The kit is intended to be used by analytical laboratories for quality control / quality assurance testing, food and agricultural testing and research purposes only (e.g. virological monitoring of production processes).

The kit is not intended for clinical diagnostics and should therefore be regarded as “For Research Use Only”.

1.1 Test Principle

After concentration of viral particles from food matrices and subsequent extraction of viral RNA the VIRSeek Food Hepatitis E Virus kit can be used for the detection of the target virus. The first step of a real-time RT-PCR is a reverse transcription (RT) of viral RNA to cDNA, which can then be amplified by real-time PCR. For the extraction of RNA we recommend the VIRSeek RNAExtractor Food kit (see section 1.3.1).

For virus concentration and extraction of soft fruits, leaf, stem and bulb vegetables and bivalve molluscan shellfish we recommend the protocol recommended by the ISO 15216-1/-2.

According to ISO guidelines 15216 the processes and horizontal methods for detection of HAV and norovirus using real-time RT-PCR in food samples require the usage of a process control virus in order to verify the correct virus concentration process from samples. Process control viruses need to have similar characteristics to the target virus of interest. Therefore the Eurofins GeneScan Technologies GmbH VIRSeek Food Murine Norovirus (MNV) Process Control kit, in combination with all food-borne virus detection kits (see section 1.3.1), is recommended. DNA amplification and detection methods take advantage of the nucleotide sequence conservation found in viral genomes that ensures the potential for high specificity and sensitivity in detection of food-borne pathogenic viruses.

By means of specific primers nucleotide sequences of the hepatitis E virus are amplified during PCR from isolated and reverse-transcribed total cDNA. Primers do not cross-react with transcribed RNA (cDNA) from other common food-borne virus species including norovirus genogroup I, norovirus genogroup II, hepatitis A virus, rotavirus, sapovirus or astrovirus and DNA derived from other relevant biological material.

1.2 Components of the Kit

For real-time RT-PCR: cat. no. 5728200501

- 1x **OligoMix HEV***, vial with amber cap with primers / probes for IPC / HEV and IPC-RNA, 530 µL, store at -20 °C ± 2 °C, do not freeze / thaw more than 3 times.
- 1x **BasicMix* VIRSeek Food**, vial with white cap, 265 µL, store at -20 °C ± 2 °C, do not freeze / thaw more than 3 times.
- 1x **Positive Control HEV**, vial with red cap, 100 µL, store at -20 °C ± 2 °C, do not freeze / thaw more than 6 times.
- 1x **Negative Control**, vial with transparent cap, 500 µL, store at -20 °C ± 2 °C.

* Aliquot during first usage if small sample numbers are frequently analysed.

1.3 Additional Equipment, Consumables and Reagents Required

Equipment:

- 1x **Stepper pipette (1 mL)**, (e.g. HandyStep® S (Brand®), Eurofins GeneScan Technologies GmbH, cat. no. 5617703401).
- 1x **Single channel pipette (1 mL, 100 µL)**, (e.g. Transferpette® S 100 - 1000 µL (Brand®), Eurofins GeneScan Technologies GmbH, cat. no. 5617703301).
- 1x **Single channel pipette (100 µL, 10 µL)**, (e.g. Transferpette® S 10 - 100 µL (Brand®), Eurofins GeneScan Technologies GmbH, cat. no. 5617703201).
- 1x **Single channel pipette (up to 10 µL)**, (e.g. Transferpette® S, 0.5 - 10 µL (Brand®), Eurofins GeneScan Technologies GmbH, cat. no. 5617703101).
- 1x **Cooling block** for 1.5 mL tubes.
- 1x **96 well cooling block**, (e.g. Blue cooling block 96 well, Eurofins GeneScan Technologies GmbH, cat. no. 5613900501).
- 1x **Vortex mixer**, (e.g. VWR Collection, cat. no. 444-2790).

Centrifuge for microtiter-plates / or -strips - depending on throughput:

- Capacity of 2x 8-well strips: (e.g. Carl Roth GmbH, Rotilabo® centrifuge with butterfly rotor, cat. no. T465.1).
- Capacity of 4x 8-well strips: (e.g. Sigma Aldrich Co LLC, MyFuge™ 12 mini centrifuge, cat. no. Z681733-1EA).
- Capacity of two times 12x 8-well strips: (e.g. Benchmark Scientific, PlateFuge™ microplate microcentrifuge, cat. no. 5613901701).

Real-time PCR Thermocycler:

- Agilent AriaMx™ with HEX™, FAM™ and Cy5™ filter set.
- Bio-Rad CFX96 Touch™ (CFX Manager™ Software / CFX Maestro™ Software).
- Bio-Rad CFX96 Touch™ Deep Well (CFX Manager™ Software / CFX Maestro™ Software).

Consumables:

- **RNase-free water** (molecular biology grade).
- **DNA- / Nuclease-free pipette tips** with filters, need to be compatible with pipettes used.
- **RNase-free reaction tubes**, 1.5 mL (e.g. DNA LoBind Tubes, Eppendorf, cat. no. 0030108051).
- **RNase-free pipette tips** need to be compatible with pipettes used.
- **PCR plates or strips**, compatible with thermocycler used.
- **Optical 8-caps strip or equivalent seals** (compatible with thermocycler used).
- **RNase decontaminating reagent** (e.g. RNase AWAY[®] Carl Roth GmbH, cat. no. A998).
- **DNA degrading agent** (e.g. Roti[®] Nucleic Acid-free, Carl Roth GmbH, cat. no. HP69).
- **Gloves**, powder free.

1.3.1 Complete Portfolio of the VIRSeek Food Solution

- **VIRSeek Food Hepatitis A Virus**, cat. no. 5728200101
 - Real-time RT-PCR kit with 48 reactions for rapid detection of hepatitis A virus (HAV) in food samples.
- **VIRSeek Food Hepatitis E Virus**, cat. no. 5728200501
 - Real-time RT-PCR kit with 48 reactions for rapid detection of hepatitis E virus (HEV) in food samples.
- **VIRSeek Food Norovirus Genogroup I**, cat. no. 5728200201
 - Real-time RT-PCR kit with 48 reactions for rapid detection of norovirus genogroup I (NoVGI) in food samples.
- **VIRSeek Food Norovirus Genogroup II**, cat. no. 5728200301
 - Real-time RT-PCR kit with 48 reactions for rapid detection of norovirus genogroup II (NoVGII) in food samples.
- **VIRSeek Food Murine Norovirus (MNV) Process Control**, cat. no. 5728200401
 - Murine norovirus spiking material (1 mL) and real-time RT-PCR kit with 48 reactions for rapid detection of murine norovirus (MNV) process control virus in food samples.
- **VIRSeek RNAExtractor Food**, cat. no. 5524400101
 - Kit for extraction of viral RNA via silica-coated magnetic beads from different food products (incl. soft fruits, leaf, stem and bulb vegetables, bottled water and bivalve molluscan shellfish). 48 x 500 µL or 96 x 250 µL sample input volume. All reagents provided.

2 HOW TO USE THIS PRODUCT

Important Notes:

- Store all reagents as indicated in section 1.2.
- Do not use the reagents beyond the expiration dates printed on the labels.
- Never store kit components in the vicinity of samples or post-PCR products.
- Extraction of viruses from food matrices such as soft fruits, leaf, stem and bulb vegetables and bivalve molluscan shellfish should be performed according to the ISO 15216 protocol.

2.1 General and Safety Precautions

- All samples should be handled with caution as they are potentially infectious.
- HEV should not be handled by pregnant women, children, elderly and immuno-compromised individuals due to the high infection risk and potentially fatal health consequences for this group, in particular for the unborn child in case of pregnant women.
- The VIRSeek Food Hepatitis E Virus kit contains no hazardous components. For more information, please refer to the VIRSeek kit SDS.
- The VIRSeek Food Hepatitis E Virus kit contains glycerol and propane-1,2-diol which may cause mild skin irritation.

2.2 Working Guidelines

- Comply with Good Laboratory Practice (refer to EN ISO 7218 standard).
- Refer to EN ISO 22174:2005 for the general requirements for the *in-vitro* amplification of nucleic acid sequences.
- Refer to EN ISO 15216-1:2017 and EN ISO 15216-2:2019 for virus extraction from food matrices such as soft fruits, leaf, stem and bulb vegetables and bivalve molluscan shellfish.
- Perform cleaning protocol (outlined in section 2.4).
- Use DNA-, nuclease-free, and sterile lab ware.
- Wear gloves and change frequently.

2.3 RNA Handling – Specific Working Guidelines

It is important to create and maintain an RNase-free environment when working with RNA. RNases are very thermostable enzymes degrading RNA – even in small quantities. Laboratory personnel are the main source for RNase contamination as RNases are expressed in human keratinocytes and are present on skin and hairs.

- Separate the different procedures spatially. Ideally use separate rooms for sample preparation and PCR setup laid-out to maintain a strict “one-way-system”, thus avoiding cross-contamination in the work stream. At least dedicate different areas, equipment and consumables for each procedure.
- Establish a working area, designated as “RNase-free”, in which only RNA work is performed. If the RNase-free working area is inside a lab with non-RNase-free working areas, clearly indicate RNase-free parts, e.g. using colour tape.
- Use dedicated RNase-free lab equipment (e.g. pipettes) for RNA-related work. Glassware has to be cleaned and decontaminated before use. For decontamination we recommend baking glassware at >200 °C for ≥4 hours.
- Only use RNase-free tips and consumables which are guaranteed to be RNase-free.
- Control high risk areas for DNA / amplicon contamination on a regular basis (swabs / PCR analysis).
- Clean the real-time RT-PCR working area as described in the cleaning protocol (see section 2.4).
- Wear disposable gloves (latex or vinyl gloves) to prevent contamination with RNases which are present on human skin. Change gloves frequently during the procedure and / or after touching skin, hair, common surfaces etc.
- Wear a lab coat to prevent contamination from clothes.
- Always thaw RNA on a cooling block and store RNA at -20 °C or below.
- Handle real-time RT-PCR enzyme mix as briefly as possible at 0 °C or above. Do not mix reagents from different kits and do not mix reagents from different batches. Return all reagents to -20 °C after usage.
- Store VIRSeek kit components for real-time RT-PCR in dedicated areas, and separate from sample storage.
- Only open one tube at a time and always change pipette tips between liquid transfers to avoid cross-contamination.

2.4 Cleaning Protocol

Before commencing work and after completing the work, ensure that the real-time RT-PCR working area is cleaned as follows:

Cleaning steps	Cleaning protocol
1.	Decontaminate surfaces with Roti [®] Nucleic Acid-free [*] or 1 % HCl to remove DNA / RNA contamination.
2.	Clean the work surfaces and non-disposable laboratory equipment (pipettes, shaker, thermo shaker etc.) with an RNase decontaminating solution [*] (e.g. RNase AWAY [®] , Carl Roth, cat. no. A998) to remove RNase contaminations.

** Follow the manufacturer's instructions.*

2.5 Waste Disposal

Dispose of any waste which is potentially contaminated with a pathogenic virus according to your internal and local regulations.

For disposal of reagents and chemicals please refer to the material safety data sheet (SDS).

2.6 Before you Begin

Store the cooling block for real-time RT-PCR at -20 °C overnight.

Extraction of viruses from different food matrices is performed according to the ISO 15216 protocol.

For RNA extraction use suitable RNA extraction kits, for optimal performance we recommend to use Eurofins GeneScan Technologies' VIRSeek RNAExtractor Food kit (see section 1.3.1).

2.7 Real-Time RT-PCR

2.7.1 Special Precautions during Real-Time RT-PCR Analysis

RT-PCR includes the reverse transcription (RT) of RNA into cDNA. RNA is a molecule which is particularly at risk of degradation due to abundant free RNases in the environment. Prior to RT, special emphasis has to be put on RNase-free environments (see section 2.3).

PCR is an exponential reaction. Therefore, after RT and amplification, the detection of single DNA targets is possible. The extreme sensitivity requires special precautions for handling and equipment. After a successful amplification, several billion amplicons are present in the reaction tube. Each of them might lead to a false positive result when contaminating sample material, i.e. by spreading as aerosols.

2.7.2 PCR Setup

Calculate required number of reactions and pipette all components (OligoMix and BasicMix) together and mix for the final reaction mix. The final real-time RT-PCR reaction mix is prepared with an additional 10 % volume.

Frequent freezing and thawing might cause inactivation of the reagents. Do not freeze / thaw kit components more than three times.

Components of final reaction mix	Amount per reaction	e.g. for 10 real-time RT-PCR reactions (+ 10 %)
BasicMix	5 µL	55 µL
OligoMix	10 µL	110 µL
Total volume	15 µL	165 µL

Before starting the practical working steps make sure you have switched on the computer, the PCR instrument and ensure the sample layout for the PCR plate is suitably documented and programmed (see below "Plate Setup").

1. Place PCR plate or strips into the 96-well cooling block which has been cooled at -20 °C.
2. Add 15 µL of final reaction mix to each test well.
3. Add 5 µL Positive Control HEV, Negative Control and negative extraction control sample to the corresponding wells.
4. Add 5 µL of each sample to the corresponding reaction well of the PCR plate.
5. Use optical caps to seal the PCR plate / strips.
6. Spin down the plate / strips in a centrifuge.
7. Transfer the PCR plate / strips to the real-time RT-PCR instrument and start the run according to the thermocycler's instructions.
8. Store samples at -20 °C or below in case PCR needs to be repeated.

Samples and Controls for Hepatitis E Virus Assay

Designation	Volume of reaction mix	Addition of
Test samples	15 µL	5 µL of sample
Positive control (C ⁺)	15 µL	5 µL of Positive Control HEV
Negative extraction control (E ⁻)	15 µL	5 µL of negative extraction control sample
Negative control (C ⁻)	15 µL	5 µL of Negative Control

Plate Setup

The following PCR plate setup is recommended by Eurofins GeneScan Technologies GmbH if samples are only analysed for HEV and the process control virus. The controls correspond to the controls recommended by ISO 15216.

	1	2	3	4	5	6	7	8	9	10	11	12
A	C ⁺	S1					(E ⁻) ¹	(S1) ¹				
B	C ⁻	Sn					(C ⁻) ¹	(Sn) ¹				
C	E ⁻						(PC ⁺) ¹					
D							(PC 10 ⁻¹) ¹					
E							(PC 10 ⁻²) ¹					
F							(PC 10 ⁻³) ¹					
G												
H												

¹ Run with process control virus real-time RT-PCR Assay (e.g. VIRSeek Food Murine Norovirus (MNV) Process Control, refer to section 1.3.1)

- C⁺: positive control (of the target of interest)
- C⁻: negative control
- E⁻: negative extraction control
- S1-Sn: test samples
- (PC⁺): process control
- (PC (10⁻¹/10⁻²/10⁻³)): process control standard (10⁻¹ / 10⁻² / 10⁻³ dilution of RNA extraction from process control sample)

Thermal Profile

1 HOLD	1 HOLD	45 CYCLES	
reverse transcription	enzyme activation & reverse transcriptase inactivation	denaturation	annealing & extension
20 min at 50 °C	3 min at 95 °C	10 sec at 95 °C	60 sec at 60 °C
no data collection	no data collection	no data collection	data collection

For Bio-Rad CFX96 Touch™ Standard and Deep Well use default ramp rate.

Probe / Detection System

VIRSeek Food Hepatitis E Virus	Fluorophore (Dye)
HEV	FAM™
IPC	Cy5™

3 DATA INTERPRETATION

Data is analysed manually by using the appropriate software provided by the cycler manufacturer.

Set baseline and threshold values according to manufacturers' instructions.

For the validated cyclers we recommend the following settings:

Real-time RT-PCR Thermocycler	Threshold	Baseline
Agilent AriaMX™	Auto ¹⁾	Adaptive
Bio-Rad CFX96 Touch™	Auto	Baseline Subtracted Curve Fit ²⁾
Bio-Rad CFX96 Touch™ Deep Well		Baseline Subtracted Curve Fit ²⁾

¹⁾ If appropriate, auto calculated threshold with default background based threshold settings can be used: Cycle range: 5 thru 9; Sigma multiplier: 10

²⁾ Always apply fluorescence drift correction

If the threshold is set too high in automatic mode, adjust it manually. For orientation the amplification curve of the positive control should be used. The threshold should be set at the beginning of the exponential phase of this curve.

3.1 Export of Raw Data

For raw data export please follow the instruction in the corresponding cyler analysis software.

3.2 Evaluation of Results

The following tables provide an overview of the criteria to evaluate the run files:

Control evaluation

Control type	HEV	IPC	Overall results
Positive control (C ⁺)	$22 \leq Cq \leq 38$	Not relevant	Valid
	$Cq < 22$		Invalid*
	$Cq > 38$		Invalid*
Negative control (C ⁻)	No Cq	$Cq \leq 37$	Valid
	No Cq	$Cq > 37$	Invalid*
	$Cq \neq \text{"No Cq"}$	Not relevant	Invalid*

*Check amplification curve for sigmoid amplification signals, software background calculation and threshold settings

Scoring of samples

Target name	Cq result	Target specific results
HEV	$15 \leq Cq \leq 43$	Positive
	$Cq < 15$	Check amplification curve for sigmoid amplification signals, software background calculation and threshold settings.
	$Cq > 43$	Compare to negative control. Check amplification curve, software background calculation and threshold settings.
	No Cq	Negative
IPC	$Cq (C^-) - 3 \leq Cq \text{ Sample} \leq Cq (C^-) + 3$	Valid
	$Cq \text{ Sample} < Cq (C^-) - 3$	Unexpected result. Check amplification curve for sigmoid amplification signals, software background calculation and threshold settings.
	$Cq \text{ Sample} > Cq (C^-) + 3$	Sample inhibited.
	No Cq	Sample inhibited.

Final result interpretation for qualitative HEV real-time RT-PCR (including process control virus)

Preliminary sample result	IPC	Process control virus	Final results	Warning / measure
Positive for HEV	Not relevant	Not relevant	Positive for HEV	
Negative for HEV	Not inhibited	Valid	Negative for HEV	
Negative for HEV	Inhibited	Valid	Sample inhibited	Test 1:10 dilution of RNA extract of undiluted sample, see also ISO 15126. If 1:10 dilution is still inhibited, repeat RNA extraction of the sample.
Negative for HEV	Inhibited	Invalid	Inhibited and extraction efficiency of process control virus is too low	Process control virus potentially inhibited, test 1:10 dilution of sample and process control virus. If 1:10 dilution is still inhibited, repeat RNA extraction of the sample. If process control virus is still invalid, repeat virus extraction.
Negative for HEV	Not inhibited	Invalid	Extraction efficiency of process control virus is too low	Repeat virus extraction

Final result interpretation for qualitative HEV real-time RT-PCR (without process control virus)

Preliminary Target Result	IPC	Final results	Warning/ measure
Positive for HEV	Not relevant	Positive for HEV	
Negative for HEV	Not inhibited	Negative for HEV	
Negative for HEV	Inhibited	Sample inhibited	Test 1:10 dilution of RNA extract of undiluted sample. If 1:10 dilution is still inhibited, repeat RNA extraction of the sample.

4 TROUBLESHOOTING

Observation:

No or weak fluorescence signal in FAM channel for the positive control

Possible Cause	Solution
Inappropriate storage of one or more kit components or kit is expired	Kit components have to be stored under appropriate conditions (see section 1.2) and only used up to the expiry date (printed on the kit label). Do not mix kit components from different lot numbers.
Kit components went through too many freeze-thaw cycles	Kit components (BasicMix, OligoMix) should not be freeze-thawed more than three times as it may cause inactivation of the reagents.
Selected channel does not conform to the protocol	Ensure that FAM channel has been chosen for analysis of the virus-specific amplification; select Cy5 channel for analysis of the internal positive control (IPC) amplification (see section 2.7.2).
Incorrect programming of the thermal profile	Check if thermal profile was defined according to the protocol (see section 2.7.2).
Incorrect real-time RT-PCR reaction mix setup	Real-time RT-PCR has to be repeated. Ensure that you use the correct volumes of OligoMix and BasicMix (see section 2.7.2).
Positive control not added to the test well	Make sure you added the positive control to the correct test well.

Observation:

No or weak Cy5 fluorescence signal of the internal control RNA (IPC)

Possible Cause	Solution
Inappropriate storage of one or more kit components, or kit components are expired	Kit components have to be stored under appropriate conditions (see section 1.2) and only used up to the expiry date (printed on the kit label).
Kit components went through many freeze-thaw cycles	Kit components (BasicMix, OligoMix) should not be freeze- thawed more than three times as it may cause inactivation of the reagents.
Real-time RT-PCR is inhibited	<p>If fluorescence signal is only missed in one or more samples, but signals are detected in all of your controls, your sample/s may contain PCR-inhibitors.</p> <p>Common PCR-inhibitors are phenol, polysaccharides, detergents and compounds from source material (plant / animal tissue), which are carried-over from the sample preparation.</p> <p>Repeat real-time RT-PCR with a 1:10 dilution of your samples to examine if the inhibitory effect can be reduced.</p> <p>Make sure that you use an appropriate RNA isolation method, which removes potential real-time RT-PCR inhibitors.</p> <p>Ethanol as a common component of wash buffers has to be completely removed at the end of your RNA isolation.</p>
Selected channel does not conform to the protocol	Cy5 channel has to be selected for analysis of the amplification of the internal positive control (IPC) (see section 2.7.2).
Incorrect programming of the thermal profile	Check if thermal profile was defined according to the protocol (see section 2.7.2).
Incorrect RT-PCR reaction mix setup	<p>Real-time RT-PCR has to be repeated.</p> <p>Ensure that you use the correct volumes of OligoMix and BasicMix (see section 2.7.2).</p>

Observation:

Fluorescence signal in the FAM channel of the negative control (C⁻)

Possible Cause	Solution
Baseline setting	Look at the amplification curve of the negative control. If a sigmoidal curve occurs, there is a potential contamination. However, if the curve is a slowly rising linear line, the auto baseline end setting of the cycler should be checked. Check the settings and adjust it manually. The amplification curve of the positive control should be used as a reference.
Contamination of your reactions	Real-time RT-PCR has to be repeated. If the C ⁻ is negative in the repetition, the contamination occurred while loading the samples into the PCR reaction tubes. Only use RNA / DNA-free lab ware and consumables. Control all areas for DNA / amplicon contamination. Always clean the working area and equipment with Roti® Nucleic Acid-free or 1 % HCl to avoid DNA contamination from previous PCRs. PCR reaction tubes have to be closed immediately after loading the samples. If negative control C ⁻ shows a fluorescence signal in the repetition, one or more of the kit components or the consumables (e.g. tubes) are contaminated. Use a new kit and observe Good Laboratory Practice (refer to EN ISO 22174 standard). We recommend using separate working areas for mix preparation, sample loading, and the real-time RT-PCR.

Observation:

No or weak fluorescence signal in FAM and / or Cy5 channel(s)

Possible Cause	Solution
Incorrect operation of the real-time PCR cycler	Check if your cycler operates according to the required specification. If necessary, perform cycler test, or contact the corresponding cycler service.

Observation:

Poor reproducibility between replicate samples

An increased variation in C_q-values across replicates can be due to poor pipetting technique or non-calibrated / imprecise pipettes.
Comply with Good Laboratory Practice (refer to EN ISO 7218 standard) and check your equipment.

5 PRODUCT WARRANTIES, SATISFACTION GUARANTEE

Eurofins GeneScan Technologies GmbH (“GeneScan”) warrants the products manufactured by it will be free of defects in materials and workmanship when used in accordance with the applicable instructions before the expiration date marked on the product packaging and when stored under the storage conditions recommended in the instructions and/or on the package. GeneScan makes no other warranty, expressed or implied. There is no warranty of merchantability or fitness for a particular purpose.

GeneScan’s sole obligation with the respect to the foregoing warranties shall be, at its option, to either replace or to refund the purchase price of the product(s) or part thereof that proves defective in materials or workmanship within the warranty period, provided the customer notifies GeneScan promptly of any such defect. GeneScan shall not be liable for any direct, indirect or consequential damages resulting from economic loss or property damages sustained by buyer or any customer from the use of the product(s). A copy of Eurofins GeneScan Technologies GmbH terms and conditions can be obtained on request, and is also provided in our price lists.

6 PRODUCT USE LIMITATIONS

This kit is developed, designed, and sold for research purposes only. It is not to be used for diagnostic purposes or analysis of food and feed unless expressly cleared for that purpose by the competent regulatory authorities in the country of use. All due care and attention should be exercised in the handling of the materials described in this text.

7 IMPORTANT NOTES

Registered names, trademarks, etc. used in this document, even when not specifically marked as such, are not to be considered unprotected by law.



TECHNICAL SUPPORT SERVICE

For technical assistance and more information please contact the Eurofins GeneScan Technologies GmbH Customer Service or your local distributor.

Eurofins GeneScan Technologies GmbH

Engesser Str. 4

79108 Freiburg, Germany

Phone: + 49-(0)761-5038-200

Fax: + 49-(0)761-5038-111

kits@eurofins.com

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