

INgezim GLUTEN

R.30.GLU.K.2

Double Antibody Sandwich (DAS) ELISA kit for the
detection of Gluten residues.

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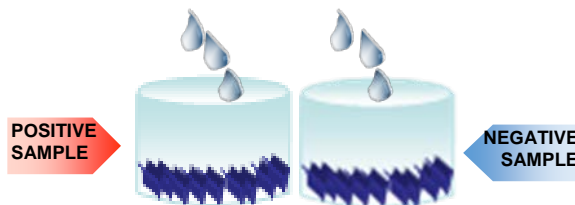
1 PRODUCT APPLICATION

The product INgezim GLUTEN has been designed for quantitative analysis of Gluten proteins in food and environmental samples.

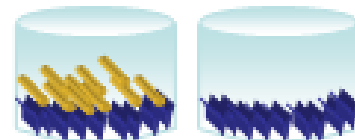
2 TECHNICAL BASIS OF THE PRODUCT

The product is based on the Double Antibody Sandwich (DAS) ELISA technique, which is summarized in the following scheme.

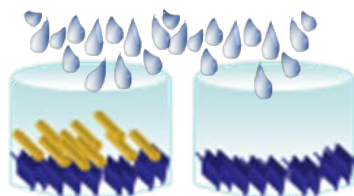
- Plates are supplied coated with R5 monoclonal antibodies specific for Prolamins. On these wells, samples are added and incubated



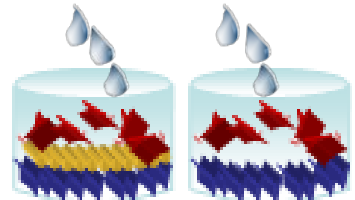
- If samples contain the antigen, it will bind the antibodies specific for Prolamins, which are coating the plate.



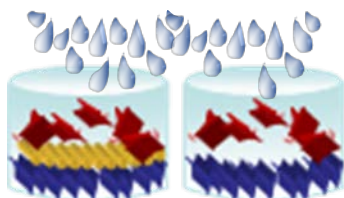
- At this point a washing step is needed to remove all non-specifically fixed materials



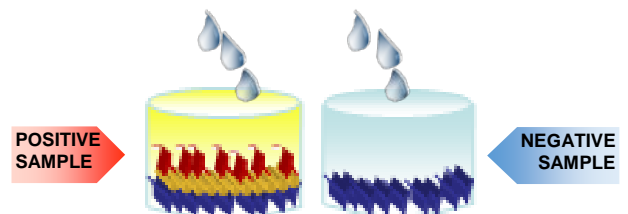
- If we add the R5 antibody specific for Gluten, it will bind to the antigen of the sample. For future detection of this antigen, this antibody is labelled with Peroxidase.



- Again a washing step is required to remove all non specifically fixed conjugate



- If a substrate for peroxidase is added, a colorimetric reaction will appear.





3 KEY REAGENTS USED

The optimum performance of the test is partially due to the key reagents used in its formulation, which, are:

- Capture Monoclonal Antibody: R5 monoclonal antibody specific for prolamins of wheat, rye and barley (no oats).
- Conjugate: R5 monoclonal antibody specific for prolamins of wheat, rye and barley (no oats) HRPO (Horseradish peroxidase) conjugated.

4 REFERENCE MATERIAL

| EUROPEAN GLIADIN FROM THE PWG | |
|-------------------------------|----------------------------------|
| Product Description | Reference gliadine (lyophilized) |
| Supplier | Prolamin Working Group |
| Available Presentations | Vial of 100 mg |
| Storage | 4°C |

5 VALIDATION PROTOCOL

The production of every new lot implies the following internal validation steps:

- a. Preparation and validation of the standard curve. The OD of every standard should meet pre-fixed values.
- b. Validation of internal and external reference samples (FAPAS). Following the kit insert, every sample should meet a pre-fixed value according with the standard points.
- c. Preparation and validation of the Conjugate, according with the correspondent standard points and Negative Control
- d. The absorbance value of the Positive Control should be higher than the absorbance of the 25ng/ml point
- e. The absorbance value of the Negative Control should be lower that the absorbance of the 1,56 ng/ml point
- f. Variability of the standard points and reference samples
- g. Intraplate variability
- h. Interplate variability



6 SENSITIVITY

6.1 LIMIT OF DETECTION AND QUANTIFICATION USING THE EUROPEAN STANDARD

In order to determine the sensitivity of the assay, the European standard was used. Five different dilutions of it were made. The results obtained showed that the assay was able to detect 1,56ng/ml of this standard. This value corresponds to an amount of **3ppm of gluten** in foods according to the next formula and supposing a sample dilution of 1/25.

$$\text{ppm} = (C \times D \times 2 \times 40) / 1000$$

Where:

C: concentration of the sample calculated from the calibration curve in ng/ml.

D: Dilution factor of the sample (25, 50, 100, etc.)

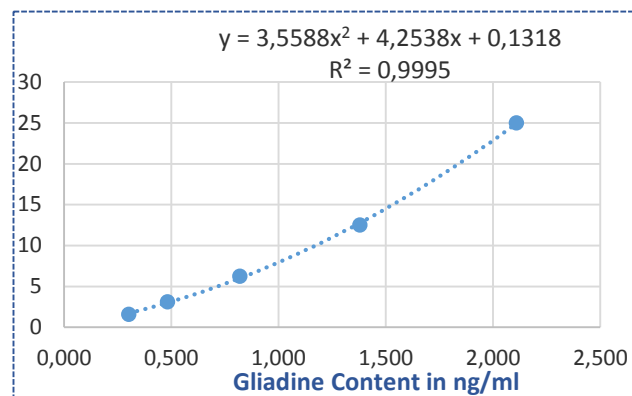
2: Applied factor to express the results in gluten concentration

40: Dilution factor used during the sample preparation (0,25g in 10 ml of extraction buffer).

1000: conversion from ng/ml to ppm

TABLE I

| STANDARDS (ng/mL) | OD (450) |
|----------------------|----------|
| 25 | 2,109 |
| 12,5 | 1,378 |
| 6,25 | 0,819 |
| 3,12 | 0,482 |
| 1,56 | 0,301 |





6.2 LIMIT OF QUANTIFICATION IN FOOD MATRIX

Five Gluten Free samples were fortified at the lowest level of quantification of the assay (3 ppm of Gluten). All samples were assayed 3 times, obtaining the results showed in the Table below (Table II)

TABLE II (Recovery rates at LOQ level)

| Matrix | Assay 1 (ppm) | Assay 2 (ppm) | Assay 3 (ppm) | Average | CV (%) |
|---------------------------|---------------|---------------|---------------|---------|--------|
| York ham | 3.2 | 3.2 | 2.4 | 2.9 | 15 |
| Baby food (with 6 fruits) | 2.6 | 4.86 | 3.04 | 3.5 | 34 |
| Soy milk | 4.9 | 3.02 | 3.52 | 3.8 | 25 |
| Gluten free cookies | 4.6 | 4.3 | 4.2 | 4.4 | 5 |
| Chocolate with milk | 2.9 | 2.3 | 2.01 | 2.4 | 19 |

The presence of gluten at the limit of quantification (3 ppm) was reliably detected in all cases; as the absorbance values of the fortified samples were higher than the negative control, blank for reagents, and the correspondent non fortified samples.

7 SPECIFICITY AND SELECTIVITY

The specificity of the R5 Monoclonal Antibody used in the assay was determined by the Unit of Gluten of the National Center of Biotechnology (CNB); by using immunoblotting technique. This R5 MAb specifically detects the prolamins of gluten present in WHEAT, RYE and BARLEY (gliadins, secalin and hordein respectively).



For checking the specificity and cross-reactivity of the assay, 50 samples (free of gluten) were extracted and analysed, following the instructions of the INGEZIM GLUTEN kit (30.GLU.K2). The results are summarized in the table below (Table III)

TABLE III (Specificity and Cross-Reactivity)

| | MATRIX | Result |
|----|--|--------------------|
| 1 | Sweetener (powder) | Negative (< 3 ppm) |
| 2 | Sweetener (tablets) | Negative (< 3 ppm) |
| 3 | Brewer's yeast | Negative (< 3 ppm) |
| 4 | Chips (maize) | Negative (< 3 ppm) |
| 5 | Snacks | Negative (< 3 ppm) |
| 6 | Snacks with cheese flavor | Negative (< 3 ppm) |
| 7 | Black chocolate (74%) | Negative (< 3 ppm) |
| 8 | White chocolate | Negative (< 3 ppm) |
| 9 | Melted chocolate | Negative (< 3 ppm) |
| 10 | Hot chocolate | Negative (< 3 ppm) |
| 11 | Mint tea | Negative (< 3 ppm) |
| 12 | Gluten free baby food (Milupa) | Negative (< 3 ppm) |
| 13 | Gluten free baby food (Nestlé) | Negative (< 3 ppm) |
| 14 | Yeast for cooking | Negative (< 3 ppm) |
| 15 | Gluten free rustic bread | Negative (< 3 ppm) |
| 16 | Gluten free baby food (dextrinated cereals) | Negative (< 3 ppm) |
| 17 | Gluten free baby food (Nutricia) | Negative (< 3 ppm) |
| 18 | Gluten free baby food (Nutriben) | Negative (< 3 ppm) |
| 19 | Custard | Negative (< 3 ppm) |
| 20 | Gelatin | Negative (< 3 ppm) |
| 21 | Mize flour | Negative (< 3 ppm) |
| 22 | Fish soup | Negative (< 3 ppm) |
| 23 | Rice crackers with chocolate | Negative (< 3 ppm) |
| 24 | Cereals mixture (rice, maize and dry fruits) | Negative (< 3 ppm) |
| 25 | Cornstarch | Negative (< 3 ppm) |
| 26 | Rice flour | Negative (< 3 ppm) |
| 27 | Rice semolina | Negative (< 3 ppm) |
| 28 | Custard with canola | Negative (< 3 ppm) |
| 29 | White chocolate | Negative (< 3 ppm) |
| 30 | Gluten free pre-cooked bread | Negative (< 3 ppm) |
| 31 | Cereals with coconut | Negative (< 3 ppm) |
| 32 | Milk chocolate | Negative (< 3 ppm) |
| 33 | Rice crackers with soy | Negative (< 3 ppm) |
| 34 | Maize crackers | Negative (< 3 ppm) |
| 35 | White chocolate with nuts | Negative (< 3 ppm) |



| | | |
|----|--------------------------------------|--------------------|
| 36 | Milk chocolate with almonds | Negative (< 3 ppm) |
| 37 | Black chocolate (60%) | Negative (< 3 ppm) |
| 38 | Quick yeast for cooking | Negative (< 3 ppm) |
| 39 | Cornflour | Negative (< 3 ppm) |
| 40 | Chicken bouillon (tablet) | Negative (< 3 ppm) |
| 41 | Meat bouillon (tablet) | Negative (< 3 ppm) |
| 42 | Fish bouillon (tablet) | Negative (< 3 ppm) |
| 43 | Sweet pepper | Negative (< 3 ppm) |
| 44 | Rice crackers with caramel | Negative (< 3 ppm) |
| 45 | Gluten free flour | Negative (< 3 ppm) |
| 46 | Cookies with chocolate (gluten free) | Negative (< 3 ppm) |
| 47 | Corn with honey | Negative (< 3 ppm) |
| 48 | Baby food with gluten free cereals | Negative (< 3 ppm) |
| 49 | Tea with lemon | Negative (< 3 ppm) |
| 50 | Crème caramel | Negative (< 3 ppm) |



8 RECOVERY DATA

8.1 RECOVERY USING REFERENCE SAMPLES

Some reference samples were analyzed by using the INGEZIM GLUTEN assay. These samples includes fortified samples (spikes), FAPAS samples and reference samples from other laboratories. Results are showed in the Table below (Table IV):

TABLE IV (RECOVERY OF REFERENCE SAMPLES)

| Sample | Theoretical Gluten content (ppm) | INGEZIM GLUTEN result (ppm) | Recovery (%) |
|----------------|----------------------------------|-----------------------------|--------------|
| Spike 0 ppm | 0 | < 1.56 | - |
| Spike 20 ppm | 20 | 24 | 120 |
| Spike 50 ppm | 50 | 58 | 116 |
| Spike 100 ppm | 100 | 114 | 114 |
| Spike 200 ppm | 200 | 196 | 98 |
| Fapas Cake Mix | 20.5 | 22 | 107 |
| Fapas 20 ppm | 20 | 22 | 110 |
| M 104 65 ppm | 65 | 54 | 83 |
| 10 ppm A+ | 10 | 12 | 120 |
| 100 ppm A+ | 100 | 105 | 105 |
| M 83 21 ppm | 21 | 25 | 119 |
| M 93 25 ppm | 25 | 28 | 112 |
| Fapas 2795A | 70 | 71 | 101 |



8.2 RECOVERY AT 20 ppm

A range of matrixes was fortified with 10 ppm of gliadin (20 ppm of Gluten). Results by using INGEZIM GLUTEN kit are showed in the table below (Table V).

TABLE V (Recovery data at 20ppm spiking level)

| SAMPLES | ppm |
|-----------------------------------|-----|
| Soy drink with cinnamon and lemon | 26 |
| Vegetables juice | 22 |
| Pineapple drink with soy | 17 |
| Orange and carrot juice | 23 |
| Tiger nut milk drink (horchata) | 24 |
| Coconut milk | 29 |
| Bolognese sauce | 19 |
| Almond drink | 23 |
| Vinaigrette | 19 |
| Ketchup | 21 |
| Barbecue sauce | 24 |
| Fried tomato | 26 |
| Mustard sauce | 30 |
| Orange juice | 23 |
| Vegetable soap | 22 |
| Rice drink | 28 |
| Soy drink with papaya and mango | 23 |
| Soy drink with chocolate | 16 |
| Balsamic vinaigrette with honey | 18 |



9 VARIABILITY

9.1 STANDARD CURVE VARIABILITY

The standards of three different kits were assayed by duplicate several times and at different days, in order to assess the absorbance variability. Results are showed in the table below:

TABLE (Standards curve absorbance at 450 nm)

| | Positive Control | 25 ng/mL | 12,5 ng/mL | 6,25 ng/mL | 3,12 ng/mL | 1,56 ng/mL | Negative Control | Blank of reagents |
|-----------------|------------------|----------|------------|------------|------------|------------|------------------|-------------------|
| Kit 1 (Assay 1) | 3.096 | 2.260 | 1.637 | 0.984 | 0.677 | 0.443 | 0.157 | 0.157 |
| Kit 1 (Assay 2) | 3.025 | 2.148 | 1.500 | 0.961 | 0.642 | 0.431 | 0.146 | NA |
| Kit 1 (Assay 3) | 3.212 | 2.575 | 1.824 | 1.216 | 0.744 | 0.514 | 0.149 | 0.138 |
| Kit 2 (Assay 1) | 3.325 | 2.421 | 1.936 | 1.246 | 0.755 | 0.514 | 0.194 | 0.166 |
| Kit 2 (Assay 2) | 2.735 | 2.184 | 1.528 | 0.974 | 0.678 | 0.480 | 0.174 | 0.167 |
| Kit 3 (Assay 1) | 3.373 | 2.546 | 1.833 | 1.138 | 0.729 | 0.488 | 0.262 | 0.137 |
| Kit 3 (Assay 2) | 3.104 | 2.292 | 1.862 | 1.106 | 0.681 | 0.396 | 0.187 | 0.148 |
| Kit 3 (Assay 3) | NA | 2.181 | 1.599 | 1.024 | 0.673 | 0.407 | 0.251 | 0.141 |
| Average | 3.124 | 2.326 | 1.715 | 1.081 | 0.697 | 0.459 | 0.190 | 0.150 |
| CV % | 6.8 | 7.2 | 9.8 | 10.4 | 5.7 | 10.1 | 23.4 | 9.2 |

The CV % for the duplicates was < 10% in all the cases. In addition, there was no significant differences comparing the absorbance values of the standards between different kits and different assays.



9.2 INTRAPLATE VARIABILITY

The same point was analysed in all the wells of the same ELISA plate.

Absorbance values and variability data are showed below:

| Intraplate variability | | | | | | | | | | | |
|------------------------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|
| 1,15 | 1,111 | 1,093 | 1,136 | 1,042 | 1,144 | 1,136 | 1,071 | 1,204 | 1,11 | 1,186 | 1,103 |
| 1,099 | 1,051 | 1,046 | 0,933 | 1,046 | 1,052 | 1,04 | 1,12 | 1,088 | 1,155 | 1,176 | 1,103 |
| 1,111 | 1,05 | 1,01 | 0,989 | 1,005 | 1,01 | 1,011 | 1,052 | 1,065 | 1,142 | 1,161 | 1,13 |
| 1,139 | 1,183 | 0,985 | 0,975 | 1 | 0,991 | 0,996 | 1,035 | 1,08 | 1,135 | 1,146 | 1,145 |
| 1,132 | 1,062 | 1,017 | 1,028 | 1,032 | 1,187 | 1,037 | 1,061 | 1,109 | 1,143 | 1,163 | 1,152 |
| 1,126 | 1,073 | 1,022 | 1,042 | 1,017 | 1,02 | 1,034 | 0,948 | 1,079 | 1,112 | 1,162 | 1,142 |
| 1,195 | 1,108 | 1,075 | 1,1 | 1,121 | 1,113 | 1,111 | 0,983 | 1,146 | 1,182 | 1,191 | 1,162 |
| 1,02 | 1,15 | 1,148 | 1,169 | 1,153 | 1,202 | 1,187 | 1,117 | 1,123 | 1,209 | 1,182 | 1,112 |

| Average | σ | CV |
|---------|----------|------|
| 1,094 | 0,066 | 6,04 |

9.3 INTERPLATE VARIABILITY

The same point was analysed in 12 strips wells, every strip from a different plate.

Absorbance values and variability data are showed below:

| Interplate variability | | | | | | | | | | | |
|------------------------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|
| Plate 1 | Plate 2 | Plate 3 | Plate 4 | Plate 5 | Plate 6 | Plate 7 | Plate 8 | Plate 9 | Plate10 | Plate11 | Plate12 |
| 1,093 | 1,046 | 0,948 | 1,041 | 0,952 | 1,051 | 1,062 | 1,098 | 1,13 | 1,117 | 1,135 | 1,076 |
| 1,024 | 1,001 | 0,921 | 0,954 | 0,996 | 0,974 | 0,984 | 0,99 | 1,021 | 1,032 | 0,961 | 1,119 |
| 1,036 | 0,934 | 0,911 | 0,901 | 0,939 | 0,943 | 0,959 | 0,947 | 1,006 | 0,996 | 1,038 | 1,119 |
| 0,913 | 0,967 | 0,908 | 0,905 | 0,922 | 0,915 | 0,928 | 0,932 | 1,012 | 1,003 | 1,059 | 1,12 |
| 1,058 | 0,961 | 0,915 | 0,923 | 0,952 | 0,965 | 0,971 | 0,987 | 1,011 | 1,044 | 1,106 | 1,079 |
| 1,024 | 0,958 | 0,914 | 0,958 | 0,933 | 0,917 | 0,948 | 0,956 | 0,965 | 1,016 | 1,118 | 1,153 |
| 1,101 | 1,042 | 1,017 | 0,939 | 1,027 | 1,017 | 0,934 | 0,971 | 1,055 | 1,128 | 0,982 | 1,124 |
| 0,962 | 1,1 | 1,06 | 1,084 | 1,061 | 1,053 | 1,102 | 1,115 | 1,1 | 1,102 | 1,107 | 1,143 |

| Average | σ | CV |
|---------|----------|------|
| 1,028 | 0,069 | 6,73 |



9.4 INTRA EXTRACTION VARIABILITY

Sample A and Sample B were extracted, one time each. Both extractions were tested 5 times each, obtaining the results showed below

| Exp # | Sample A | |
|---------------|----------|--------------|
| | OD | Gluten (ppm) |
| N° 1 | 1,01 | 10,0 |
| N°2 | 0,98 | 13,3 |
| N°3 | 1,11 | 12,9 |
| N°4 | 1,14 | 11,3 |
| N°5 | 1,00 | 10,4 |
| Mean | 1,05 | 11,60 |
| Standard Dev. | 0,07 | 1,31 |
| VC* | 6,79 | 11,37 |

| Exp # | SampleB | |
|---------------|---------|--------------|
| | OD | Gluten (ppm) |
| N° 1 | 0,895 | 1,9 |
| N°2 | 0,785 | 2,1 |
| N°3 | 0,989 | 2,6 |
| N°4 | 0,979 | 2,1 |
| N°5 | 0,851 | 1,9 |
| Mean | 0,90 | 2,10 |
| Standard Dev. | 0,09 | 0,25 |
| VC* | 9,59 | 12,08 |

9.5 INTER EXTRACTION VARIABILITY

Samples A and B were extracted at 5 different days. Obtained results are showed below:

| Exp # | SampleA | |
|---------------|---------|--------------|
| | OD | Gluten (ppm) |
| N° 1 | 1,00 | 9,7 |
| N°2 | 0,97 | 13,1 |
| N°3 | 0,98 | 10,0 |
| N°4 | 1,03 | 9,2 |
| N°5 | 0,89 | 8,0 |
| Mean | 0,97 | 10,00 |
| Standard Dev. | 0,05 | 1,69 |
| VC* | 5,31 | 16,90 |

| Exp # | Sample B | |
|---------------|----------|--------------|
| | OD | Gluten (ppm) |
| N° 1 | 0,908 | 1,9 |
| N°2 | 0,86 | 2,6 |
| N°3 | 0,918 | 2,2 |
| N°4 | 0,95 | 1,9 |
| N°5 | 0,82 | 1,7 |
| Mean | 0,89 | 2,06 |
| Standard Dev. | 0,05 | 0,31 |
| VC* | 5,75 | 15,20 |



9.6 INTERBATCH VARIABILITY USING REFERENCE SAMPLES

Reference samples with a known gluten content, were analysed by using 3 different lots of INGEZIM GLUTEN kits.

Average results, standard deviation and CV are showed in the table below:

| Reference Sample | Batch A | Batch B | Batch C | Average | σ | CV |
|--------------------|---------|---------|---------|---------|----------|------|
| Sample 1 (10 ppm) | 11 | 9 | 9 | 9,6 | 1,1 | 11,9 |
| Sample 2 (20 ppm) | 21 | 19 | 23 | 21 | 2 | 9,5 |
| Sample 3 (100 ppm) | 99 | 93 | 100 | 97 | 3,7 | 3,8 |

10 PRODUCT LIMITATION

The assay is not designed for being used with samples where the GLUTEN proteins have been hydrolysed by any kind of sample treatment.

The most common hydrolysed samples are beer, whiskey, some syrups and some baby food.

For this kind of samples, INGEZIM HYDROLYSED GLUTEN (30.GLH.K2) assay should be used.



11 FAPAS PROFICIENCY TESTING RESULTS

The performance of the INgezim Gluten assay (30.GLU.K2) has been tested by participating in some proficiency testing (FAPAS); and obtaining equivalent results compared with other kits from the market (including our INgezim Gluten Quick kit 30.GL2.K2)

The assigned value (x_a) was determined for each analyte where sufficient data allowed and, in conjunction with the standard deviation for proficiency (σ_p) was used to calculate a z-score for each result.

Results for the proficiency tests are summarized in the tables below:

Proficiency test 27227

| | Assigned value, x_a mg/mL | Number of scores, $ z \leq 2$ | Total number or scores | % $ z \leq 2$ |
|--|-----------------------------|--------------------------------|------------------------|----------------|
| Ingenasa INgezim Gluten (30.GLU.K2) | 52.0 | 10 | 12 | 83 |
| Ingenasa INgezim Gluten Quick (30.GL2.K2) | 56.0 | 7 | 7 | 100 |
| Neogen Veratox for Gliadin R5 (8510) | 37.3 | 14 | 16 | 88 |
| R-Biopharm Ridascreen Gliadin (R7001) | 50.0 | 90 | 97 | 93 |
| R-Biopharm Ridascreen Fast Gliadin (R7002) | 46.2 | 6 | 6 | 100 |
| Romer Labs AgraQuant ELISA Gluten G12 (COKAL02000) | 50.7 | 3 | 3 | 100 |

Proficiency test 27228

| | Assigned value, x_a mg/mL | Number of scores, $ z \leq 2$ | Total number or scores | % $ z \leq 2$ |
|---|-----------------------------|--------------------------------|------------------------|----------------|
| Ingenasa INgezim Gluten (30.GLU.K2) | 54.0 | 9 | 9 | 100 |
| Ingenasa INgezim Gluten Quick (30.GL2.K2) | 56.0 | 6 | 6 | 100 |
| Neogen Veratox for Gliadin R5 (8510) | 30.0 | 9 | 9 | 100 |
| R-Biopharm Ridascreen Gliadin (R7001) | 45.0 | 52 | 58 | 90 |