DN*Animal Ident* Goose

IPC (LR/HR)

Cat. No. 5422220810

Test kit for the qualitative detection of goose DNA with IPC, 96 real-time PCR reactions

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15. TECHNICAL SERVICE

If you have any questions or experience any difficulties regarding this kit or GeneScan Technologies products in general, please do not hesitate to contact us. GeneScan customers are a major source of information regarding advanced or specialised uses of our products. This information is helpful to other scientists as well as to the researchers at GeneScan. We therefore encourage you to contact us if you have any suggestions about product performance or new applications and techniques. For technical assistance and more information please call the GeneScan Technologies Technical Service Department or your local distributor.

Eurofins GeneScan Technologies GmbH

Engesser Str. 4
79108 Freiburg, Germany
Phone: + 49-(0)761-5038-100
Fax: + 49-(0)761-5038-111
kits@eurofins.com
www.eurofins.com/kits

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Test kits, their components and instructions for use are subject to alterations. They are intended for research purposes only.

DN*Animal Ident* Goose IPC Kit

Kit for qualitative real-time PCR detection of goose DNA

1. INTRODUCTION

National regulations in most countries of the world require declaration of components of food and feed and a corresponding labelling.

Declaration of components of animal origin are crucial e.g. for vegetarian food, food for religious communities, export and trade.

The Eurofins GeneScan DN*Animal* kits comprise several major advantages:

- High sensitivity (down to 0.01% admixture)
- High specificity (primers and probe)
- Robust test methods
- Fast results

The DN*Animal Ident* Goose IPC kit was designed for a general detection of goose (*anser*) DNA in DNA extracted i.e. from food and feed matrices.

The complete test comprises the following steps:

1. DNA extraction (not included in this kit)
2. Real-time PCR detection of the target

Animal species can be analysed in most matrices, even in gelatine. The processing steps commonly used in food and feed production usually reduce the fragment length of the sample DNA, but DNA is not completely degraded and the base sequence of the fragments is not changed. However, extremely rigid (acid/heat) processing can degrade DNA in a way that amplification is impossible.

Specific DNA sequences are amplified by RT-PCR (Real-time Polymerase Chain Reaction) and detected with high specificity due to the Taqman™ probes.

The probe of the detection system in this kit uses FAM® as reporter dye and the IPC (internal positive control) uses R6G. The probes use non-fluorescent quenchers. The kit is validated for use on Applied Biosystems® 7500, ABI7500 Fast, Agilent MX3000P™, MX3005PTM, Agilent AriaMx™, Biorad CFX96 Touch™ or Roche LightCycler®480 and ABI 7900HT™.

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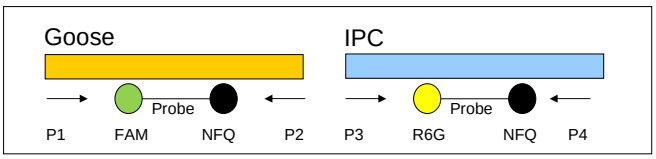
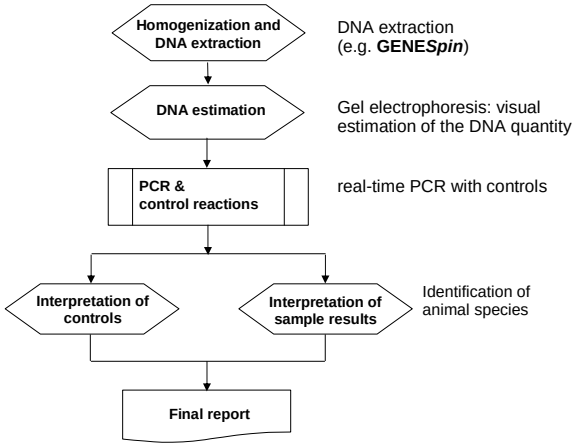


FAM™ and R6G are the fluorescent reporter dyes attached to the 5' ends of the probes for the goose target and for the IPC (internal positive control). Non-fluorescent quenchers are used for quenching.

2. TEST PROCEDURE – FLOWCHART



3. TIME SCHEDULE

Step	Hands-on time	Time involved
DNA-extraction	0.5 h	1.5 h
PCR-reaction	0.5 h	3.5 h
Total time	1.0 h	5.0 h

The hands-on time has been calculated for a single sample. When several samples are handled simultaneously, the time increases.

4. COMPONENTS OF THE KIT

The kit contains all components to run control and specific reactions for a total of 100 PCR reactions.

2x **MasterMix QL RT IPC (LR/HR+) GS-P-08.032, Goose**

1 ml composed of

- 650 µL BasicMix QL RT (NR) GS-P-26.001 • EFGi 2x (NR) 1.5 U
- 390 µL OligoMix QL RT IPC (LR/HR+) • GS-P-08.032 • Goose

Mix prior to use!

2 x **Positive control DNA**: **Genomic DNA from goose** (50 µL, 10 copies/µL)

1x **DNA stabilization buffer II**, 150 µL (for NTCs)

2x **ROX™ dye,** 23 µL (to be used only for High ROX cyclers, see instructions below)

Important Note: Store all following components at -20°C. Never store solutions and materials for DNA extraction together with samples or amplicons. Never use them in areas where PCR products (amplicons) may be present.

5. PREPARATION OF THE KIT'S REAGENTS

Store the kit at –20°C until opened for the first time.
Thaw reagents just before use, mix them by vortexing and centrifuge briefly before use.
Just thaw as many BasicMix and OligoMix vials as necessary. Frequent freezing and thawing might cause inactivation of the reagents. If smaller volumes are needed, aliquot reagents at first use.
Refer to the reagent label for specific instructions regarding the correct storage.

6. MATERIAL AND EQUIPMENT NOT INCLUDED IN THE KIT

– water, DNase-free
– Vortex
– Micropipettes (variable 1-1000 µL) and filter tips
– PCR optical tubes or plates 0.2 ml and optical caps or seals
– RT-PCR Thermocycler

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7. SAMPLE PREPARATION

7.1 DNA extraction

The extraction of DNA from the sample is a crucial step, and in order to guarantee optimal quantity and purity, an appropriate method has to be chosen for each sample matrix. For most products, the DNA can be extracted with a kit from our DNA*Extractor* kit line or with our GENE*Spin* kit.
Absolute DNA amount and purity (meaning: free of PCR inhibiting substances) affect overall sensitivity of the analysis. Low DNA amount subjected to analysis results in poor LOD with regard to the specific test sample. Insufficiently pure DNA displaying inhibitory effects may even make testing impossible.
It is recommended to use the sample DNA undiluted. In In case inhibitors are present in the DNA, dilution of the sample DNA is feasible. However, it needs to be considered that the practical/sample LOD for the individual analysis of the sample will be affected and – in case the DNA amount used is too small – may not reach the method LOD, which means that the sensitivity for the test is decreased. DNA amount can be determined prior to analysis by one of the following methods:
- Real-time PCR monitor run (preferred method)

- Fluorimetric or spectrophotometric or other physical measurements, e.g. gel electrophoresis may be applied.

Especially the spectrophotometric methods can suffer significant errors due to interfering DNA impurities or partial DNA denaturation/degradation.
For samples known with regard to extraction yield and purity, DNA measurement may be omitted.

Each sample should be treated in duplicate.

It is recommended to perform an extraction control for each set of samples extracted simultaneously, i.e. a complete DNA extraction without sample material, which should subsequently undergo PCR analysis.

8. PCR

8.1 Special precautions during PCR

PCR is an exponential reaction. Detection of single target DNA copies is possible. The extreme sensitivity requires special precautions for handling and equipment. A successful amplification leads to several billion amplicons in the reaction tube. Each of them might lead to a false positive result when contaminating sample material or PCR, e.g. by spreading in aerosols.
Most important rules to avoid false-positive results are:

a) Separate the different procedures spatially. Use separate rooms for sample preparation, amplification and analysis of amplicons and dedicate separate equipment and materials for each procedure. Mixing of PCR components should be done in a separate room, best on a clean bench. Keep all working steps separate from rooms where gel electrophoresis of amplicons is performed.
b) Use filter tips for micropipettes.
c) Wear disposable powder-free gloves.
d) Never store GMO*Ident* Kits and materials for DNA extraction together with samples or amplicons.
e) Always perform extraction controls and PCR controls (NTCs).

8.2 Preparation of MasterMix

Calculate the number of reactions and amount of equivalent MasterMix volume before thawing and mixing the reagents and starting the practical work.
The MasterMix is made of pre-made reagents ready for direct use after mixing of the two components.

Please be aware that the NTCs and positive controls (C+) are indispensable for the evaluation of the test and must be included in every run.
PCR is performed in a volume of 25 µL in 0.2 mL reaction tubes/plates according to the RT-PCR cycler instructions.

20 µL MasterMix for low ROX (LR) cyclers consist of 12.5 µL BasicMix + 7.5 µL OligoMix per reaction.

If you use one of the following cyclers, add also more ROX™dye: ABI 5700, 7000, 7300, 7700, 7900, Opticon 2 or other cyclers laid out for high ROX (HR) concentration (please enquire in case of doubt).
ROX™ is added increasing the volume of the MasterMix to 20.45 µL.
20.45 µL HR MasterMix consist of 12.5 µL BasicMix + 0.45 µL ROX (50x) + 7.5 µL OligoMix per reaction.

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The following reactions are required for a RT-PCR run with 1, 8, 23 or 46 samples (incl. extraction controls). Add approx. 5% of volume as pipetting error compensation or use complete tubes.

Samples	1	8	23	46
NTCs	2	2	2	2
Pos. controls	2	2	2	2
Samples (duplicate)	2	16	46	92
Total no. reactions	**6**	**20**	**50**	**96**
Total MM volume	**120 µL**	**400 µL**	**1 mL**	**2 mL**
BasicMix	75 µL	250 µL	625 µL or 1 tube	1250 µL or 2 tubes
OligoMix	45 µL	150 µL	375 µL or 1 tube	750 µL or 2 tubes
ROX (only for HR!)	*2.7 µL*	*9 µL*	*22.5 µL*	*43.2 µL*

1. Thaw only the volumes of BasicMix and OligoMix needed for analysis. Shake thoroughly.
2. Remove the required volumes and mix in a fresh tube. Freeze the rest.
3. If the cycler requires a high ROX™ level, add ROX™.
4. Mix thoroughly.
5. The composed MasterMix can be stored up to 4 h in the refrigerator, but must not be used longer.

8.3 PCR Setup

Every run requires two no template controls (NTC) and two positive controls (C+). It is highly recommended to test the samples in duplicate.

1. Label all PCR reaction tubes.
2. Add 20 µL of the composed MasterMix to the wells.
3. Add 5 µL of stabilisation buffer to NTCs.
4. Add 5 µL of control (goose) DNA to positive controls.
5. Add 5 µL of sample DNA to test reactions.

	1	2	3	4	5	6	7	8	9	10	11	12
A	NTC	NTC	C+	C+	1a	1b	2a	2b	3a	3b	4a	4b
B	5a	5b	6a	6b	7a	7b	8a	8b	9a	9b	10a	10b
C	11a	11b	12a	12b	13a	13b	14a	14b	15a	15b	16a	16b
D	17a	17b	18a	18b	19a	19b	20a	20b	21a	21b	22a	22b
E	23a	23b	24a	24b	25a	25b	26a	26b	27a	27b	28a	28b
F	29a	29b	30a	30b	31a	31b	32a	32b	33a	33b	34a	34b
G	35a	35b	36a	36b	37a	37b	38a	38b	39a	39b	40a	40b
H	41a	41b	42a	42b	43a	43b	44a	44b	45a	45b	46a	46b

Plate layout for 46 samples.
NTC = no template control;
C+ = positive control;
a and b = sample duplicates

8.4 Programming of Plate Documents

Before starting the practical work, program the plate document und the cycling conditions.

For description of the instrument programming please refer to the user manual of the respective instrument and software version.
For further setup and evaluation information for your individual cycler, please see our application notes for your cycler model on our website www.eurofins.com/kits.

Program your template with the following settings:

Agilent Mx3005P™ (3000P™)

System	Assay	Filter
Goose	8032a	FAM™/SYBR® Green I
IPC	8032i	HEX™
Reference	ROX	ROX™

The fluorescence signals are scanned in the following order: (1) ROX, (2) FAM, (3) HEX.

The filter set gain settings for ROX, FAM and HEX are: pmt = 1x

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ABI 7500 (7500 Fast) / ABI 7900 HT™

System	Detector	Reporter	Quencher
Goose	8032a	FAM™	NONE
IPC	8032i	VIC™	NONE

Passive Reference: ROX

- ABI 7500 Fast
  PCR is performed in the “Standard 7500” run mode.

- ABI 7900HT
  Activate 9600 emulation mode.

Agilent AriaMx™

System	Target Name	Dye
Goose	8032a	FAM™
IPC	8032i	HEX™
Reference	ROX	ROX™

Bio-Rad CFX96 Touch™

System	Targets	Dyes
Goose	8032a	FAM™
IPC	8032i	VIC™

Default maximum Ramp rate is used.
A reference dye is not used.

Cycling Conditions

The run parameters are to be programmed as follows:

Thermal Cycler Times and Temperatures		
1 HOLD	**CYCLE (45 repeats)**	
enzyme act.	*denaturation*	*ann. & extension*
10 min at 95°C	**15 sec at 95°C**	**90 sec at 60°C**
no data collection	no data coll.	data collection

For other thermocyclers than the mentioned ones, it may be necessary to optimise the PCR parameters.

Roche LightCycler®480:
Color Compensation for FAM, R6G performed according to the manual. . (cat. # 5427200201)
FAM signal is collected using the FAM detector
R6G signal is collected using the VIC/HEX /Yellow555 detector
Ramp rate is set to 4.4 for heating and 2.2 for cooling.
A reference dye is not used

Save the created file/document.

For more setup and evaluation information for your individual cycler, please send an email to kits@eurofins.com or see our website www.eurofins.com/kits.

9. RESULTS

9.1 Evaluation

Refer to your cycler´s manual for details. An evaluation (Excel™) sheet can be requested at kits@eurofins.com.

Threshold:
ABI 7500/7500 Fast /7900HT™ Agilent Mx3005P™/3000P™:
The threshold should be placed in the region of exponential amplification across all of the amplification plots. This region is depicted in the log view of the amplification plots as the portion of the plot, which is linear. The threshold line should neither be placed in the plateau phase nor in the initial linear phase of amplification, which may be too low and into the background fluorescence or where splitting between replicates may be observed.

Agilent AriaMx™:
Auto calculated threshold with default background based threshold settings can be used when appropriate:
Cycle range: 5 thru 9
Sigma multiplier: 10

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Bio-Rad CFX96 Touch™:
Auto calculated threshold can be used but should be checked visually.
Baseline:

ABI 7500/7500 (Fast), ABI7900 HT™:
Set manually, 3-15 or auto baseline settings

Agilent Mx3005/3000P™: Adaptive

Roche LightCycler®480:
Choose “Abs Quant/2nd Derivative Max” and High Sensitivity settings to obtain CP values.
Choose “Endpoint Genotyping” analysis mode to obtain Endpoint Fluorescence.

Agilent AriaMx™: Adaptive

Bio-Rad CFX96 Touch™:
Baseline Subtracted Curve Fit and Fluorescence Drift Correction should be applied.

9.2 Interpretation of Results

Export Ct values to the Excel™ sheet provided (please request to kits@eurofins.com), or create an own evaluation sheet following the parameters below, where you will find an overview of the algorithms applied for scoring of the results in our Excel™ Evaluation Sheet.

Definitions

Threshold cycle (Ct/Cp/Cq) Cut-offs	goose Cut-off-1: Mean Ct (C+) -5 IPC Cut-off-1: Mean Ct (NTC) -3
	goose Cut-off-2: Mean Ct (C+) +8 IPC Cut-off-2: Mean Ct (NTC) +3
Endpoint fluorescence (EPF)-Limits	goose: Mean EPF (C+) x 0.2 IPC: Mean EPF (NTC) x 0.33
Outliers	Maximum acceptable outliers (C+): 0 Maximum acceptable outliers (NTC): 0

Inhibition control, scoring of IPC

Threshold cycle (Ct/Cp/Cq) = Ct and Endpoint fluorescence = EPF	**Results**
Ct_{IPC} Cut-off-1 ≤ Ct_{IPC} sample ≤ Ct_{IPC} Cut-off-2 **and** EPF_{IPC} sample ≥ EPF_{IPC} Limit	Sample valid
Ct_{IPC} sample < Ct_{IPC} Cut-off-1 **and** EPF_{IPC} sample ≥ EPF_{IPC} Limit	Check amplification!
Ct_{IPC} sample > Ct_{IPC} Cut-off-2 **or** EPF_{IPC} sample < EPF_{IPC} Limit	Sample invalid
No Ct_{IPC}	Sample invalid

Test reaction (goose)

Threshold cycle (Ct/Cp/Cq) =Ct	**Endpoint fluorescence dRN/dR/EF= EPF**	**Result**
Ct Cut-off-1 ≤ Ct sample ≤ Ct Cut-off-2	EPF sample ≥ EPF sample Limit	Reaction positive
Ct sample ≤ Ct Cut-off-2	EPF sample < EPF sample Limit	Check amplification!
Ct sample < Ct Cut-off-1	EPF sample ≥ EPF sample Limit	Check amplification!
Ct sample > Ct Cut-off-2	EPF sample ≥ EPF Limit	Reaction positive
Ct sample > Ct Cut-off-2	EPF sample < EPF Limit	Reaction negative
No Ct	-	Reaction negative
***Note*:** In case of a Ct value 40.0 (for LC480) or "Check amplification!" the linear scale amplification plots must be carefully checked for presence or absence of a sigmoid PCR amplification signal. If a sigmoid curve can be observed, the sample is positive; if not, the sample is negative.		

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Final result from combination of inhibition control and test reaction

IPC	**goose test**	**Final result**
Sample valid	Reaction positive	Positive
Sample valid	Reaction negative	Negative
Sample invalid	Reaction positive	Positive
Sample invalid	Reaction negative	Inhibited

Evaluation of the IPC
Calculate the MEAN Ct (= Ct/Cp/Cq) value from NTC. Refer to data from IPC-detector. To calculate the Ct cut-off-1, subtract 3 Ct, to calculate the Cut-off-2 add 3 Ct. Calculate the MEAN Endpoint fluorescence (EPF) value from NTC. Refer to data from IPC-detector. The Endpoint fluorescence cut-off is 33% of the MEAN Endpoint fluorescence.

Evaluation of goose specific test
Calculate the MEAN Ct/Cp/Cq value from positive Control. Refer to data from goose detector, respectively. To calculate the Ct/Cp/Cq cut-off-1, subtract 5 Ct/Cp/Cq, to calculate the Cut-off-2, add 8 Ct/Cp/Cq. Calculate the MEAN Endpoint fluorescence value from positive control. Refer to data from goose detector, respectively.

The Endpoint fluorescence cut-off is 20% of the MEAN Endpoint fluorescence.
Combine results of IPC and the goose test to the final result.

9.3 Ambiguous results

If independently extracted DNAs show deviations in the results for "A" and "B", this may be due to sample material non-homogeneity or to non-uniformities in the DNA-extraction efficiency. Repeat DNA extraction and homogenize sample more thoroughly. If the ambiguous result persists, it is most likely due to a concentration close to the LOD.

10. LIMIT OF DETECTION

The relative limit of detection has been validated at 0.01% of goose DNA in a total DNA content of 150 ng/reaction (goose DNA in chicken DNA, w/w)
The absolute detection limit of the method is ≤ 10 copies per reaction.
The limit of detection for the analysis of individual samples depends on several factors: the presence of PCR-inhibiting substances, the capability of the DNA extraction method to remove these inhibitors and the

degree of DNA damage, and thus strongly on the type of sample and the DNA extraction procedure. Quantification is not possible with this kit.

11. PRODUCT USE LIMITATIONS

The GeneScan DN*Animal* Kit is developed, designed, and sold for research purposes only. It is not to be used for diagnostic purposes or analysis of food and feed unless expressly cleared for that purpose by the appropriate regulatory authorities in the country of use. All due care and attention should be exercised in the handling of many of the materials described in this text.

12. PRODUCT WARRANTIES AND SATISFACTION GUARANTEE

Eurofins GeneScan Technologies GmbH ("GeneScan") warrants the products manufactured by it will be free of defects in materials and workmanship when used in accordance with the applicable instructions and before the expiration date marked on the product packaging under the storage conditions, recommended in the instructions and/or on the package. Application protocols published by GeneScan are intended to be only guidelines for the buyers of the products. Each buyer is

expected to validate the applicability of each application protocol to his individual application. GeneScan makes no other warranty, expressed or implied. There is no warranty of merchantability or fitness for a particular purpose.
GeneScan's sole obligation with the respect to the foregoing warranties shall be, at its option, to either replace or to refund the purchase price of the product(s) or part thereof that proves defective in materials or workmanship within the warranty period, provided the customer notifies GeneScan promptly of any such defect. GeneScan shall not be liable for any direct, indirect or consequential damages resulting from economic loss or property damages sustained by buyer or any customer from the use of the product(s). A copy of Eurofins GeneScan Technologies GmbH terms and conditions can be obtained on request and is provided in our price lists.

13. IMPORTANT NOTES

- The TaqMan processes are covered by patents issued to Hoffmann-La Roche AG and Applied Biosystems Inc., which are applicable in certain countries. Eurofins GeneScan does not encourage or support the unauthorized or unlicensed use of these processes.

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Use of this kit is recommended for persons that either have a license or are not required to obtain a license.

- Registered names, trademarks, etc. used in this document, even when not specifically marked as such, are not to be considered unprotected by law.

14. TROUBLESHOOTING

Result	Possible mistakes/reasons	Possible verification and measures
No PCR signals from samples	Inhibition of PCR by inhibitory substances. Inhibition by too much DNA.	Clean DNA further* or dilute DNA solution. Check DNA concentration/dilution.
No PCR signals from positive controls	Wrong PCR program.	Check and correct PCR program.
No amplification, neither from control DNA nor from sample DNA	MasterMix not properly prepared Wrong PCR program.	Prepare fresh MasterMix, repeat PCR. Check program.
Positive PCR result for NTC	Contamination with DNA/amplicons when mixing the PCR components.	Optimise your precautions. Check your solutions. Decontaminate your equipment. Repeat the PCR.
Positive PCR result for extraction control	Contamination with sample material/DNA/ amplicons/ during DNA extraction or PCR setup.	Check your solutions. Repeat extraction and PCR.

* Repeat extraction of DNA from the sample. Repeat washing with 75 % ethanol three times. If necessary, clean DNA further e.g. with our DNA Cleaning Columns (cat. no. 5224700310).