

# CERTIFICATION

## **AOAC<sup>®</sup>** *Performance Tested*<sup>SM</sup>

Certificate No. **052005** 

The AOAC Research Institute hereby certifies the method known as:

### **SENSISpec INgezim Gluten R5**

manufactured by

EUROFINS INGENASA Av. De la Institución Libre de Enseñanza, 39 28037 MADRID, SPAIN

This method has been evaluated in the AOAC<sup>®</sup> *Performance Tested Methods*<sup>SM</sup> Program and found to perform as stated by the manufacturer contingent to the comments contained in the manuscript. This certificate means that an AOAC<sup>®</sup> Certification Mark License Agreement has been executed which authorizes the manufacturer to display the AOAC *Performance Tested* <sup>SM</sup> certification mark along with the statement - "THIS METHOD'S PERFORMANCE WAS REVIEWED BY AOAC RESEARCH INSTITUTE AND WAS FOUND TO PERFORM TO THE MANUFACTURER'S SPECIFICATIONS" - on the above-mentioned method for a period of one calendar year from the date of this certificate (September 09, 2021 – December 31, 2022). Renewal may be granted at the end of one year under the rules stated in the licensing agreement.

Scott Coates

Scott Coates, Senior Director Signature for AOAC Research Institute September 09, 2021

Date

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METHOD NAME(S) SENSISpec INgenzim Gluten R5	CATALOG NUMBER 30.GLU.K2			
INDEPENDENT LABORATORY Q Laboratories, Inc. 1930 Radcliff Drive Cincinnati, Ohio USA	AOAC EXPERTS AND PEER REVIEWERS Terry Koerner <sup>1</sup> , Joe Boison <sup>2</sup> <sup>1</sup> Health Canada, Ontario, CANADA <sup>2</sup> EJ Consultancy, Saskatoon, Canada			
APPLICABILITY OF METHOD Target Analyte(s) – Prolamin proteins from Gluten: gliadins from wheat, secalins from rye, and hordeins from barley	REFERENCE METHOD AOAC OMA 2012.01 (2)			
Matrixes – (0.25 g): gluten-free bead mix, oat flour stainless steel (10 cm x 10 cm)				
Performance claims - Detection and quantification of gliadin gluten-free bread and oats. Qualitative detection of gliadin on stainless steel surfaces.				
ORIGINAL CERTIFICATION DATE May 22, 2020	CERTIFICATION RENEWAL RECORD Renewed annually through December 2022.			
METHOD MODIFICATION RECORD NONE	SUMMARY OF MODIFICATION NONE			
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#### PRINCIPLE OF THE METHOD (1)

The method is based on a double antibody sandwich enzyme immunoassay using the R5 monoclonal antibody (mAb), which recognizes an epitope common to the prolamin fraction from wheat (gliadins), rye (secalins) and barley (hordeins). The assay involves the following steps:

- 1) The sample is ground (if required) and dissolved in solution buffer to bring relevant target molecules into solution.
- 2) An aliquot of 100  $\mathbb{Z}L$  is then pipetted into the wells of the ELISA plate containing the bound -specific R5 antibodies. If gluten is contained in the sample, it will be bound by the antibodies on the plate.
- 3) Using washing buffer, the remains of the unbound material is washed out.
- 4) Add a volume of 100 μL of the R5 mAb conjugated with Horse-radish peroxidase (HRPO) to each well. This conjugated mAb will bind to the gluten previously captured by the coated antibody.
- 5) Using washing buffer, the unbound mAb conjugate is washed out.
- 6) 100 μL of TMB (Tetra-methylbenzidine) is added as a substrate of the HRPO enzyme (change of the colorless substrate solution into a blue product).
- 7) A stop Solution is added, to transform the blue color into yellow one. The intensity of yellow color can be measured with an ELISA reader (at 450 nm).
- 8) The gliadin content of the samples is determined by interpolation of their OD in the calibration curve. The calibration curve is established using the standard of the Prolamin Working Group (PWG). The obtained ng/mL gliadin value is converted to gluten content by using the formula explained in the calculations sheet.

#### **DISCUSSION OF THE VALIDATION STUDY (1)**

After the rigorous validation study to confirm the performance claims of the INGENASA SensiSpec INgezim Gluten R5 Kit, all the results obtained were satisfactory according with the expected. More in detail, the linearity study analyzing five replicate test portions of the curve shows a very high reproducibility. The food matrix study demonstrates no differences, depending on the previous treatment of the sample, gliadin recovery after spiking the samples.

The recovery from the incurred bakery matrix samples tested was also as expected (98.4%). The cross-reactivity/interference study demonstrates that the assay does not produce false positive results with the selected matrixes and there are no interferences with them. As a final comment, we can indicate that the assay is valid for the detection of gluten in contaminated surfaces.

In addition, the data from the independent validation study support the product claims of the INGENASA SensiSpec INgezim Gluten R5 method in fresh raw select foods and stainless steel surfaces.

#### ERUOFINS INGENASA SENSISpec Ingenzim Gluten R5, AOAC® Performance Tested<sup>5M</sup> certification number 052005

Table 5. Results of the test of the samples from Matrix study by the INGENASA and reference test. The table includes the information about: the mean of the mean of gliadin per kg of product obtained as a mean of the different replicates, the standard deviation, the coefficient of variation (CV or RDSr), the Bias and the % of recovery. (1)

Sample	Contamination mg/kg Gliadin	Method	Mean	Standard Deviation	RDSr	Bias	% Recovery
	0	INGENASA	<loq< th=""><th>NA</th><th>NA</th><th>NA</th><th>NA</th></loq<>	NA	NA	NA	NA
Gluten free bread		REFERENCE	<loq< th=""><th>NA</th><th>NA</th><th>NA</th><th>NA</th></loq<>	NA	NA	NA	NA
	1.70	INGENASA	1.60	0.41	25.63	-0.10	94.40
		REFERENCE	<loq< th=""><th>NA</th><th>NA</th><th>NA</th><th>NA</th></loq<>	NA	NA	NA	NA
	2.00	INGENASA	1.70	0.42	24.71	-0.33	83.30
		REFERENCE	<loq< th=""><th>NA</th><th>NA</th><th>NA</th><th>NA</th></loq<>	NA	NA	NA	NA
	3.12	INGENASA	2.80	0.56	20.00	-0.30	90.40
mix		REFERENCE	3.00	1.03	34.33	-0.12	96.15
	6.25	INGENASA	5.78	0.95	16.40	-0.47	92.00
		REFERENCE	7.43	0.23	3.10	1.18	119
	12.50	INGENASA	11.11	0.09	0.80	-1.39	89.00
		REFERENCE	12.19	0.21	1.80	-0.31	98.00
	20.00	INGENASA	17.90	2.24	12.51	-2.10	89.50
		REFERENCE	24.20	0.70	2.89	4.20	121.00
	0	INGENASA	<loq< th=""><th>NA</th><th>NA</th><th>NA</th><th>NA</th></loq<>	NA	NA	NA	NA
		REFERENCE	<loq< th=""><th>NA</th><th>NA</th><th>NA</th><th>NA</th></loq<>	NA	NA	NA	NA
	1.70	INGENASA	1.68	0.15	0.09	-0.02	99.12
		REFERENCE	<loq< th=""><th>NA</th><th>NA</th><th>NA</th><th>NA</th></loq<>	NA	NA	NA	NA
	2.00	INGENASA	2.13	0.16	0.07	0.13	106.66
		REFERENCE	<loq< th=""><th>NA</th><th>NA</th><th>NA</th><th>NA</th></loq<>	NA	NA	NA	NA
Oat	3.12	INGENASA	3.62	0.46	0.13	0.50	116.15
		REFERENCE	3.60	0.52	14.44	0.48	115.38
	6.25	INGENASA	6.88	0.17	2.50	0.63	110.00
	13.50	REFERENCE	7.65	0.05	0.60	1.40	122.00
	12.50	INGENASA	13.01	0.15	1.20	0.51	104.00
		REFERENCE	16.66	1.80	10.80	4.16	133.00
	20.00	INGENASA	20.60	1.68	0.08	0.60	103.00
		REFERENCE	23.00	0.50	2.10	3.00	115.00
White rice flour	100.00	INGENASA	98.20	3.06	3.10	-1.78	98.20
White rice flour	100.00	INGENASA	108.03	9.20	8.50	8.00	108.03
White rice flour	100.00	INGENASA	103.00	2.05	2.00	3.84	103.00
White rice flour	100.00	INGENASA	98.60	1.90	1.90	-1.40	98.60
White rice flour	100.00	INGENASA	95.70	2.80	2.90	-4.30	95.70

Table 7. Summary of the results obtained with the incurred gluten-free bread samples. The table shows extractions from 1 to 10 that are analyzed by the candidate method (Sensispec INgezim Gluten R5) and extractions from 11 to 13 that are analyzed by the reference method. In the table is included the Optical density (OD at 450nm) obtained, the ppm of gliadin, the mean of OD of the different extractions, the standard deviation and de % of recovery. \*NA: Not applicable (1)

Test	Contamination	Extraction	OD 450 nm	ppm gliadin	Mean	Std dev σ	% Recovery
		Extr 1	0.113	<loq< th=""><th></th><th></th><th></th></loq<>			
		Extr 2	0.131	<loq< th=""><th rowspan="5">NA</th><th rowspan="4"></th><th rowspan="9">NA</th></loq<>	NA		NA
		Extr 3	0.139	<loq< th=""></loq<>			
		Extr 4	0.107	<loq< th=""></loq<>			
Sensispec		Extr 5	0.125	<loq< th=""></loq<>			
INgezim Gluten R5		Extr 6	0.097	<loq< th=""><th>NA</th></loq<>		NA	
	0ppm	Extr 7	0.102	<loq< th=""><th></th><th rowspan="3"></th></loq<>			
		Extr 8	0.099	<loq< th=""><th rowspan="2"></th></loq<>			
		Extr 9	0.109	<loq< th=""></loq<>			
		Extr 10	0.16	<loq< th=""><th></th><th></th></loq<>			
Reference		Extr 11	0.143	<loq< th=""><th>NA</th><th rowspan="2">NA</th><th rowspan="2">NA</th></loq<>	NA	NA	NA
Method OMA	Method OMA	Extr 12	0.375	<loq< th=""><th></th></loq<>			
2012.01		Extr 13	0.139	<loq< th=""><th></th><th></th><th></th></loq<>			
		Extr 1	1.219	5.55	(		98.40
		Extr 2	1.181	5.28			
		Extr 3	1.098	4.73			
		Extr 4	1.142	5.02	4.92 0		
Sensispec INgezim Gluten R5 5 ppm		Extr 5	1.158	5.13		0.41	
		Extr 6	1.029	4.29		0.41	
	5 ppm	Extr 7	1.081	4.62			
		Extr 8	1.078	4.60			
		Extr 9	1.198	5.40			
		Extr 10	1.081	4.62			
Reference		Extr 11	0.868	5.89	6.14	0.38	122.80
Method OMA		Extr 12	0.944	6.59			
2012.01		Extr 13	0.876	5.96			

#### Table 10. The table shows the results of the test of the samples from the surfaces. $^{a}N = Number of test portions (1)$

-N = Number of test portions (1)								
Matrix	Allergen	N <sup>a</sup>	Contamination level	1:12.5 Dilution				
				Xp	PODc <sup>c</sup>	95% CI		
Stainless Steel Environmental Surfaces	Gluten	5	0 μg/mL/100cm <sup>2</sup>	0	0.00	0.00, 0.43		
		30	0.4 μg/mL/100cm <sup>2</sup>	15	0.5	0.33, 0.66		
		5	2 μg/mL/100cm²	5	1.00	0.57, 1.00		

<sup>a</sup>N = Number of test portions

<sup>b</sup>x = Number of positive test portions

<sup>c</sup>POD<sub>C</sub> = 1:12.5 Dilution positive outcomes divided by the total number of trials

#### **REFERENCES CITED**

1. Romero, C., Venteo, A., González, I., Hevia, E., Rebollo, B., SensiPeck iNgezim Gluten R5 (30.GLU.K2), AOAC® Performance Tested<sup>5M</sup> certification number 052005.

2. AOAC OMA 2012.01